

S.No.	Details of Publications
1	<b>Manimekalai, R.</b> *, Selvi, A., Jini Narayanan, Ram Vannish, R. Shalini, Gayathri, S. Rabisha, V.P. 2023. Comparative physiological and transcriptome analysis in cultivated and wild sugarcane species in response to hydrogen peroxide-induced oxidative stress. <b>BMC Genomics.</b> Doi 10.1186/s12864-023-09218-3
2	O'Connell A, Deo J, Deomano E, Wei X, Jackson P, Aitken KS, <b>Manimekalai</b> R, Mohanraj K, Hemaprabha G, Ram B, Viswanathan R, Lakshmanan P. 2022. Combining genomic selection with genome-wide association analysis identified a large-effect QTL and improved selection for red rot resistance in sugarcane. <b>Front Plant Sci.</b> 13:1021182. doi: 10.3389/fpls.2022.1021182. PMID: 36388469; PMCID: PMC9660812.
3	M Naveenarani, P Vignesh, C Mahadevaiah, R Valarmathi, R <b>Manimekalai</b> , G Hemaprabha, C Appunu. 2022. Genome-Wide In Silico Identification, Structural Analysis, Promoter Analysis, and Expression Profiling of PHT Gene Family in Sugarcane Root under Salinity Stress. <b>Sustainability</b> 14 (23), 15893
4	<b>Manimekalai</b> , R., Gayathri, S. and Singaravelu, B. (2022). Sugarcane Transcriptomics in Response to Abiotic and Biotic Stresses: A Review. <i>Sugar Tech</i> (2022). <a href="https://doi.org/10.1007/s12355-021-01098-9">https://doi.org/10.1007/s12355-021-01098-9</a>
5	Narayanan, J., Manimekalai, R., Selvi, A. Gomathi, R. And Arun, R. 2022. Physiological, Biochemical and Molecular Responses to Oxidative Stress in Saccharum spontaneum. <i>Sugar Tech</i> (2022). <a href="https://doi.org/10.1007/s12355-022-01189-1">https://doi.org/10.1007/s12355-022-01189-1</a>
6	Srikanth, J., Mahesh, P., <b>Manimekalai</b> , R., Suresha, G. S., Singaravelu, B., & Salin, K. P. (2022). Web mite Schizotetranychus krungthepensis on sugarcane in India: molecular evidence for occurrence and the way forward. <b>CURRENT SCIENCE</b> , 123(8), 1038.
7	Mintu R. Meena, Chinnaswamy Appunu, R. Arun Kumar, R. <b>Manimekalai</b> , S. Vasantha, Gopal Reddy, Ravinder Kumar, S. K. Pandey and G Hemaprabha. 2022. Recent Advances in Sugarcane Genomics, Physiology, and Phenomics for Superior Agronomic Traits. <b>Frontiers in Genetics</b> . 10.3389/fgene.2022.854936
8	Smita Nair and <b>Manimekalai</b> . R. (2021). Phytoplasma diseases of plants: Molecular diagnostics and way forward", <i>World Journal of Microbiology and Biotechnology</i> . DOI 10.1007/s11274-021-03061-y.
9.	Selvi, A., Devi, K., <b>Manimekalai</b> , R., Prathima, P. T., Valiyaparambth, R., & Lakshmi, K. (2021). High-throughput miRNA deep sequencing in response to drought stress in sugarcane. <i>3Biotech</i> , 11(7), 312. <a href="https://doi.org/10.1007/s13205-021-02857-x">https://doi.org/10.1007/s13205-021-02857-x</a>
10	<b>R. Manimekalai*</b> , Gayathri, S., Hemaprabha, G., and Selvi, A. Kandalam, M. (2020). Role of NGS and SNP Genotyping methods in Sugarcane improvement programs. <b>Critical reviews in Biotechnology</b> . DOI: 10.1080/07388551. 2020. 1765730
11	Selvi, A., Devi, K., <b>Manimekalai</b> , R., Prathima, P.T. (2020). Comparative analysis of drought-responsive transcriptomes of sugarcane genotypes with differential tolerance to drought. <i>3 Biotech</i> . DOI: 10.1007/s13205-020-02226-0
12	Taloh, A., Raju, D.V.S., Banyal, N., Kumar, G., Panda, P., <b>Manimekalai</b> , R., Carmine Marcone, C., and Rao, G.P. (2020). Genetic diversity of phytoplasma strains infecting chrysanthemum varieties in India and their possible natural reservoirs. <i>3 Biotech</i> 10.411. <a href="https://doi.org/10.1007/s13205-020-02407-x">https://doi.org/10.1007/s13205-020-02407-x</a>
13	Gayathri S, Arockiyajainmary M, Shalini R, Vannish MR, Selvi A, <b>Manimekalai</b> R. (2019). Identification of single nucleotide polymorphisms (SNPs) in the transcriptome of sugarcane variety Co 86032 exposed to oxidative stress. <i>J sugarcane Res.</i> 9:71–85
14	Devi, K., Prathima, P.T., Gomathi, R., <b>Manimekalai</b> R., Lakshmi, K., and Selvi, A. 2019. Gene expression profiling in sugarcane genotypes during drought stress and rehydration Sugar Tech. <a href="https://doi.org/10.1007/s12355-018-0687-y">https://doi.org/10.1007/s12355-018-0687-y</a>
15	<b>Manimekalai</b> , R.*, Smita Nair, A. Naganeeswaran, Anitha Karun, Suresh Malhotra, and V. Hubbali. 2018. Transcriptome sequencing and de novo assembly in arecanut, <i>Areca catechu</i> L elucidates the secondary metabolite pathway genes. <i>Biotechnology Reports</i> . 17: 63–69. doi: 10.1016/j.btre.2017.12.005. ISSN: 2215-017X
16	Devi, K., Gomathi, R. Arun Kumar, R., <b>Manimekalai</b> , R., and Selvi, A. Field tolerance and

	recovery potential of sugarcane varieties subjected to drought. 2018. Indian Journal of Plant Physiology, DOI: 10.1007/s40502-018-0367-7
17	<b>R. Manimekalai*</b> ,Jini Narayanan, R. Ranjini, M. Gokul, A. Selvi, PradheepKumar and R. Gomathi. 2018. Hydrogen Peroxide-Induced Oxidative Stress in Sugarcane and Response Expression Pattern of Stress-Responsive Gene Through Quantitative RT-PCR. (2018). Sugar Tech. <a href="https://doi.org/10.1007/s12355-018-0604-4">https://doi.org/10.1007/s12355-018-0604-4</a>
18	<b>Manimekalai, R.*</b> ,Jini, N., Gokul, M., Selvi, A., Gomathi, R., Arun Kumar. 2018. Biochemical and physiological response to oxidative stress in cultivated sugarcane and wild genera. Indian Journal of Plant Physiology. DOI :10.1007/s40502-018-0368-6
19	<b>Manimekalai, R.*</b> ,Jini Narayanan Gokul, M., A. Selvi, Arun Meena, R. Gomathi, Bakshi Ram (2017). Genome wide analysis of NAC gene family ‘sequences’ in sugarcane and its comparative phylogenetic relationship with rice, sorghum, maize and Arabidopsis for prediction of stress associated NAC genes. Agrigene, 3: 1–11. DOI: 10.1016/j.aggene.2016.10.003 ISSN: 2352-2151
20	GP Rao, TV Madhupriya, <b>R Manimekalai</b> , AK Tiwari, A Yadav. 2017. A century progress of research on phytoplasma diseases in India. PhytopathogenicMollicutes 7 (1), 1-3
21	Madhupriya, N Banyal, R Dantuluri, VS, <b>Manimekalai, R.*</b> , GP Rao . 2017. Association of different groups of phytoplasma in flower malformation, phyllody, foliar yellowing, and little leaf disease of rose ( <i>Rosa sp.</i> )The Journal of Horticultural Science and Biotechnology 92 (4), 424-431
22	Smita Nair, <b>Manimekalai, R.*</b> . GangaRaj, K.P. and Hegde, V.2016.Loop mediated isothermal amplification (LAMP) assay for detection of coconut root wilt disease and arecanut yellow leaf disease phytoplasma. World Journal of Microbiology and Biotechnology. DOI: 10.1007/s11274-016-2078-4
23	SmitaNair, <b>R. Manimekalai*</b> , V. P. Soumya and K. C. Likitha. 2016. Dual labeled probe based real time PCR method for detection of 16SrXI-B sub-group phytoplasma associated with coconut root wilt disease in India. Australasian Plant Pathology. DOI 10.1007/s13313-016-0406-7
24	<b>Manimekalai R*</b> . Soumya, V.P., Smita Nair, Rajeev, G. and Rao, G.P. 2016. PCR detection of phytoplasma from different tissues of root (wilt) diseased coconut palms.Phytopathogenic Mollicutes.6: 23-28
25	R Gopinath, KP Deesma, VP Soumya, C Mohanan, <b>Manimekalai Ramaswamy*</b> . 2016. Application of light microscopic staining techniques for the detection of phytoplasmas in yellow leaf disease affected arecanut palms in India. PhytopathogenicMollicutes 6 (2), 77-81
26	K.P Manju, <b>R Manimekalai*</b> , SA Naganeeswaran, V Arunachalam, A Karun. 2016. Microsatellites mining in date palm ( <i>Phoenix dactylifera'L.</i> ) and their cross transferability across' Arecaceae' family. Plant Omics 9 (3), 191
27	<b>Manimekalai, R*</b> .Soumya V.P.,Srinivasan, N. and Rao G.P. 2015. Comparison of 16S rDNA sequence among the phytoplasmas associated with wilt disease of coconut from different locations. Phytopathogenic Mollicutes 5(1):37-41
28	<b>Manimekalai , R*</b> .Smita Nair, K.P. Gangaraj V.P. Soumya G.P. Rao. 2015. Isolation and characterization of partial secA gene from coconut root wilt and arecanut yellow leaf disease phytoplasma and assessment of its possible use for sero diagnosis. Indian Phytopathology. 68 (4) : 438-443
29	<b>Manimekalai* R</b> ,Roshna O.M., GangaRaj K.P., Viswanathan R., Rao, G.P. 2015. ABC Transporter from Sugarcane Grassy Shoot Phytoplasma: Gene Sequencing and Sequence Characterization. Sugar Tech. DOI 10.1007/s12355-015-0396-8
30	Sumi, K., Madhupriya , Ramaswamy <b>Manimekalai, R.</b> , Rao, G.P., and SambasivaRao K.R.S. 2015. Identification and genetic relationship among phytoplasma strains infecting coconut, arecanut and oil palm in South India. Indian Phytopath. 68 (2) : 207-214 (2015)
31	Smita Nair, <b>Manimekalai, R*</b> ,Soumya VP and Rao GP. 2015. Taqman quantitative PCR for detection of Indian arecanut yellow leaf disease phytoplasma. PhytopathogenicMollicutes 5 (2): 113-116. doi: 10.5958/2249- 4677.2015.00070.5
32	<b>Manimekalai, R*</b> ,Soumya V.P., Srinivasan, N. and Rao G.P. 2015. Comparison of 16S

	rDNA sequence among the phytoplasmas associated with wilt disease of coconut from different locations. <i>PhytopathogenicMollicutes</i> 5(1):37-41 doi : 10.5958/2249-4677.2015.00061.4
33	Naganeeswaran S., ElainApshara S. <b>Manimekalai R.</b> AmalVasu and Malhotra S.K. 2015. Cocoa EST database: Comprehensive database of Cocoa Expressed Sequence Tags (ESTs) International Journal of Innovative Research in Computer and Communication Engineering. DOI: 10.15680/IJIRCCE.2015. 0311011 .ISSN(Online): 2320-9801 ISSN (Print): 2320-9798
34.	Naganeeswaran SA, <b>Manimekalai, R*</b> .ElainApshara, ManjuKP and Karun A. 2014. Standalone EST Microsatellite mining and Analysis Tool (SEMAT): For automated EST-SSR analysis in plants. <i>Tree Genetics and Genomes</i> . DOI: 10.1007/s11295-014-0785-2
35.	<b>Manimekalai, R*</b> , N.C Anil Kumar, O.M Roshna and K. Satyamoorthy. 2014. Isolation and comparative analysis of potc gene of ABC-transporter system from coconut and sugarcane - 16srXI group phytoplasmas. <i>Journal of Plant Pathology</i> . 96 (1), 35-42. (Doi: 10.4454/JPP.V96I1.003)
36.	<b>Manimekalai, R*</b> ,Smita Nair and V. P. Soumya. 2014. Evidence of 16SrXI group phytoplasma DNA in embryos of root wilt diseased coconut palms. <i>Australasian Plant Pathology</i> .43: 93-96.
37.	Smita Nair, Roshna, O.M., Soumya V.P., VinayakaHegde, Suresh Kumar, M. <b>Manimekalai R.*</b> and George V Thomas. 2014. Real-time PCR technique for Detection of Arecaut Yellow leaf Disease Phytoplasma. <i>Australasian Plant Pathology</i> .DOI: 10.1007/s13313-014-0278-7.
38	<b>Manimekalai, R*</b> ,Roshna OM., Gangaraj KP., Smita Nair, Viswanathan, R. and Rabindran, R. 2014. Molecular characterization based on spermidine/putrescine ABC transporter gene of sugarcane grassy shoot (16SrXI), coconut root wilt (16SrXI), aster yellows (16SrI) and brinjal little leaf (16SrVI) phytoplasmas. <i>PhytopathogenicMollicutes</i> . 4 (1): 16-21
39	Chaithra, M, Madhupriya, Kumar, S, , <b>Manimekalai, R.</b> and Rao, G.P. 2014. Detection and characterization of 16SrI-B phytoplasmas associated with yellow leaf disease of arecanut palm in India. <i>PhytopathogenicMollicutes</i> . 4(2), 77-82
40	<b>R. Manimekalai*</b> ,Soumya V. P., Smita Nair, George V. Thomas and V. K. Baranwal. 2014. Molecular characterization identifies 16SrXI-B group phytoplasma ('CandidatusPhytoplasmaoryzae'-related strain) associated with root wilt disease of coconut in India. <i>Scientia Horticulturae</i> 165: 288–294.
41	<b>R. Manimekalai*</b> and K.P. Manju. (2013) Prediction of microsatellites and the annotation of repeat sequences in partial coconut genome. <i>International Journal of Innovative Horticulture</i> . 2(2):117-121
42.	<b>Manimekalai R*</b> ,Smita Nair, Soumya and G. V. Thomas. 2013. Phylogenetic analysis identifies 'CandidatusPhytoplasmaoryzae'-related strain associated with Yellow Leaf Disease of Areca palm (Areca catechu L.) in India. <i>International Journal of Systematic and Evolutionary Microbiology</i> . 63, 1376–1382.
43	DilipK.Ghosh, SumitBhose, V. J. Shivankar, <b>R.Manimekalai</b> and S. Gowda. 2012. Molecular detection of <i>CandidatusPhytoplasma</i> spp. causing witches' broom disease of acid lime ( <i>Citrus aurantifolia</i> ) in India. <i>Journal of Plant Biochemistry and Biotechnology</i> . DOI 10.1007/s13562-012-0160-z
44	<b>Manimekalai, R.*</b> ,K. P. Deeshma, K. P. Manju, V. P. Soumya, Smitha Nair and K. S. Ananda. 2012. Molecular marker based genetic variability among Yellow Leaf Disease (YLD) resistant and susceptible arecanut palm (Areca catechu. L). <i>Indian Journal of Horticulture</i> , 69(4): 455-461
45	Naganeeswaran, S Elain, A, <b>Manimekalai, R.*</b> 2012. Analysis of expressed sequence tags (ESTs) from cocoa ( <i>Theobroma cacao</i> L) upon infection with <i>Phytophthora megakarya</i> . <i>Bioinformation</i> : 8 (2): 65-69. ISSN 0973-2063
46.	<b>Manimekalai R*</b> . 2012. Inter Simple Sequence Repeat (ISSR) markers for identification of germplasm and diversity analysis in coconut ( <i>Cocosnucifera</i> L.). <i>International Journal of Innovative Horticulture</i> . 1(1):63-67, 2012
47	Pradeep Kumar, S., <b>Manimekalai, R.</b> and Ranjitha Kumar, B.D. 2011. Microsatellite marker based characterization of South Pacific coconut ( <i>Cocosnucifera</i> L.) accessions. <i>International Journal of Plant Breeding and Genetics</i> 5:34 .ISSN 1819-3595

48	Sugio A, MacLean AM, Kingdom HN, Grieve VM, <b>Manimekalai R</b> , Hogenhout* SA. 2011. Diverse targets of phytoplasma effectors: from plant development to defense against insects. Annual Review Phytopathology; 49:175-95
49	<b>Manimekalai, R*</b> . Smita Nair, Soumya, V. P Roshna O. M. and Thomas, GV. 2011. Development of Real Time PCR for the detection of coconut root (wilt) phytoplasma. Current science, 101:1209-1213
50	<b>Manimekalai, R*</b> ., Anitha Karun and Sajini K.K. (2011). Polypeptide variability among the coconut zygotic embryos in two culture conditions. Progressive Horticulture. 43(1): 81- 82
51	Manju, KP, <b>Manimekalai, R*</b> . and Arunachalam, V. 2011. Microsatellites in palm (Arecaceae) sequences. Bioinformation 7 (7): 347- 351
52	<b>Manimekalai, R*</b> . P. Anoop Raj, O. M. Roshna, Anil Paul and George V. Thomas. 2011. Updated Version of PHYTO DB, the First Data Warehousing and Mining Web Server for Phytoplasma Research. Communications in Computer and Information Science, 168, (4), 548-550, DOI: 10.1007/978-3-642-22606-9_58 (springer)
53	Nuruzzaman, M., <b>Manimekalai, R.</b> , Sharoni, A.M., Satoh K., Kondoh, H., Ooka, H., Kikuchi S*. 2010. Genome-wide analysis of NAC transcription factor family in rice Gene 465:30-44.
54.	<b>Manimekalai, R*</b> . Soumya, V.P., Sathish Kumar, R., Selvarajan, R., Krishna Reddy, M., Sasikala, George V Thomas., Rajeev M and Baranwal, V.K. 2010. Molecular detection of 16Sr XI group Phytoplasma associated with Root (Wilt) Disease of coconut ( <i>Cocosnucifera L.</i> ) in India. Plant disease 94:636.
55	<b>Manimekalai, R*</b> . Sathish Kumar, P., Soumya V.P. and George V Thomas. 2010. Molecular detection of phytoplasma associated with Yellow Leaf Disease in areca palms ( <i>Areca catechu L.</i> ) in India. Plant Disease 94:1376
56	<b>Manimekalai, R*</b> . and Nagarajan, P. 2010. Bulk Line Analysis in coconut ( <i>Cocosnucifera L.</i> ) for inferring relationship between talls, dwarfs and Niu Leka Dwarf forms. Indian J. Plant Genet. Resour. 23(1): 77-81
57	<b>Manimekalai, R*</b> . and Nagarajan, P. 2010. SSR and ISSR markers based population genetic structure of coconut ( <i>Cocosnucifera L.</i> ) germplasm accessions. Indian J. Plant Genet. Resour. 23(1): 87-92.
58	<b>Manimekalai, R*</b> . Anoop, Rakesh and Thomas GV. 2009. Putative gene database of cocoa and oil palm. Journal of Plant. Crops. 39(2): 311-318
59	<b>Manimekalai, R*</b> . and Nagarajan. 2007. Use of simple sequence repeat markers for estimation of genetic diversity in coconut ( <i>Cocosnucifera L.</i> ) germplasm accessions. Journal of Plant Biochemistry and Biotechnology, 16:29-33.
60	39. Jiji, G., Karun, A., <b>Manimekalai, R.</b> , Rajesh, M.K. and Remya, P. 2007. Identification of RAPD markers linked to sex determination in palmyrah ( <i>Borassusflabellifer L.</i> ). Current Science 93: 1075-1077
61	<b>Manimekalai, R*</b> . Nagarajan, Karun, A., Parthasarathy, V.A. 2007. RAPD as useful marker for coconut ( <i>Cocosnucifera L.</i> ) DNA analysis. Indian Journal of Horticulture 64(1):1-4
62	<b>Manimekalai, R*</b> ., and Nagarajan, P. 2006. Assessing genetic relationships among coconut ( <i>Cocosnucifera L.</i> ) accessions using Inter Simple Sequence Repeat Markers. Scientia Horticulturae 108: 49-56
63	<b>Manimekalai, R*</b> . and Nagarajan, P. 2006. Inter-relationship among coconut ( <i>Cocosnucifera L.</i> ) germplasm accessions using RAPD technique. Genetic Resource Crop Evolution 53:1137-1144
64	<b>Manimekalai, R*</b> . Rubavathi, V.B., Reji J. Thomas, Nair, R.V. and Rajesh, M.K. 2006. Analysis of segregating populations of West Coast Tall variety of coconut using microsatellite markers. Journal of Plant. Crops 34:587-590
65	Anuradha, S., Chandrasekar, C. and <b>Manimekalai, R.</b> 2006. Optimal PCR parameters for RAPD analysis in <i>Theobroma cacao</i> and <i>Cola nitida</i> . Plant cell Biotech molecular biology: 7:81
66	Devakumar, K., Jayadev, K., Rajesh, M.K., Chandrasekar, A., <b>Manimekalai, R.</b> , Kumaran, P.M. and Parthasarathy, V.A. 2006. Assessment of the genetic diversity of Indian coconut accessions and their relationship to other cultivars using microsatellite markers. Plant Genetic Resource Newsletter: 145: 38

67	<b>Manimekalai, R.,</b> Nagarajan, P. and Kumaran, P.M. 2006. Comparison of effectiveness of RAPD, ISSR and SSR markers for analysis of coconut ( <i>Cocosnucifera L.</i> ) germplasm accessions. <i>Trop. Agri. Res.</i> 18: 217-226
68	<b>Manimekalai, R.,</b> Nagarajan, P., Bharathi, M., Karun, A., Kumar, S.N. and Kumaran., P.M. 2005. Genetic variation of selected progeny lines of coconut ( <i>CocosnuciferaL.</i> ) based on Simple Sequence Repeat markers. <i>Trop. Agri. Res.</i> 17 : 58-66
69	Parthasarathy, U., Parthasarathy, V.A., <b>Manimekalai, R.,</b> Niral, V. Anuradha, U. and Kumaran, P.M. 2005. Molecular diversity in coconut eco types of coastal and inland riverine ecosystems. Indian Journal of Horticulture. 62: 315-318.
70	Rajagopal, V., <b>Manimekalai, R*</b> .et al. 2005. A database for coconut crop improvement. Bioinformation1(2):75-77
71	<b>Manimekalai, R*</b> . 2005. Informative Inter Simple Sequence Repeat (ISSR) primers for genetic analysis of coconut ( <i>Cocosnucifera L.</i> ). CORD 22 (2): 40
72	Upadhyay*, A., Jeyadev, K., <b>Manimekalai, R.</b> and Parthasarathy, V.A. 2004. Genetic relationship and diversity in Indian coconut accessions based on RAPD markers. <i>Scientia Horticulturae</i> 99: 353-362
73	P Geethalakshmi, AUpadhyay, K Jayadev, R <b>Manimekalai</b> , V. Niral and V.A Parthasarathy. 2004. Genetic finger printing in coconut cultivars. <i>Asian Journal of Plant Sciences</i> 4 (6), 353-362. ISSN: 16823974, 18125697
74	<b>Manimekalai, R*</b> . Jeyadev, K., Upadhyay, A., Devakumar, K., Rajesh, M.K., Kumaran, P.M., Rajagopal, V. and Parthasarathy, V.A. 2004. Optimization of DNA amplification fingerprinng (DAF) protocol in coconut. <i>Journal of Plant. Crops</i> 32: 1-6
75	<b>Manimekalai, R*</b> . Nagarajan, P., Bharathi, M. and Naresh Kumar, S. 2004. DNA polymorphism among coconut ( <i>Cocosnucifera</i> ) cultivars and reciprocal cross derivatives. <i>Journal of Plant. Crops</i> , 32: 117 – 124
76	Anuradha, S., Upadhyay, A., <b>Manimekalai, R.</b> and Parthasarathy, V.A. 2003. An efficient method of DNA extraction from <i>Theobroma cacao</i> and <i>Cola nitida</i> leaves. <i>Agrotropica</i> , 15: 127-130